Investigation of Antiparasitic Effect of Juniperus communis L. Fruits Extracts

Juniperus communis Meyve Ekstrelerinin Antiparaziter Etkisinin Araştırılması

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ABSTRACT

Objective: *Juniperus communis (J. communis)* (common juniper) is a plant that has been used for medicinal purposes for centuries. This study aims to evaluate the antiparasitic effects of ethanol, methanol, chloroform, and water extracts of *J. communis* fruits against *Plasmodium falciparum*, *Leishmania tropica*, *Trichomonas vaginalis*, and *Blastocystis*.

Methods: The antiparasitic activities of fruit extracts prepared at room temperature using the shaking maceration method were tested against *Plasmodium falciparum* using the ring stage survival test, and against *Leishmania tropica, Trichomonas vaginalis*, and *Blastocystis* using the broth microdilution method.

Results: The chloroform extract of *J. communis* fruits was found to be effective on *Plasmodium falciparum*, *Leishmania tropica*, *Trichomonas vaginalis*, and *Blastocystis* parasites at concentrations of 15, 10, 30 and 30 μg/mL, respectively.

Conclusion: The chloroform extract of *J. communis* fruits has shown strong antiparasitic activity against the investigated parasite species. These findings support the plant's antiparasitic potential and hold promise for future medical applications. Especially its effectiveness against metronidazole-resistant *Trichomonas vaginalis* strains is important for the development of alternative treatment options. This study highlights the potential use of *J. communis* as a medicinal plant and will contribute to the literature on research related to the isolation and structural determination of its active compounds

Keywords: Antiparasitic, Juniperus communis, Blastocystis, Juniperus communis, Leishmania, Plasmodium, Trichomonas

ÖZ

Amaç: Juniperus communis (J. communis) (adi ardıç), tibbi amaçlarla uzun süredir kullanılan bir bitkidir. Bu çalışma, J. communis meyvelerinden hazırlanan etanol, metanol, kloroform ve su ekstrelerinin *Plasmodium falciparum*, *Leishmania tropica*, *Trichomonas vaginalis* ve *Blastocystis* üzerindeki antiparaziter etkilerini değerlendirmeyi amaçlamaktadır.

Yöntemler: Çalkalamalı maserasyon yöntemi oda sıcaklığında hazırlanan meyve ekstrelerinin antiparazitik aktiviteleri, Plasmodium falciparum'a karşı ring stage survival testi ile Leishmania tropica, Trichomonas vaginalis ve Blastocystis'e karşı aktiviteleri ise sıvı mikrodilüsyon yöntemi ile araştırılmıştır. Her bir parazit türü üzerinde ekstrelerin farklı konsantrasyonlarda etkinlikleri test edilmiştir.

Bulgular: *J. communis* meyvelerinin kloroform ekstresinin, *Plasmodium falciparum*'a karşı 15 μg/mL, *Leishmania tropica*'ya karşı 10 μg/mL, *Trichomonas vaginalis*'e karşı 30 μg/mL ve *Blastocystis*'e karşı ise 30 μg/mL konsantrasyonda etkili olduğu gözlenmiştir. **Sonuç:** *J. communis* meyvelerinin kloroform ekstresi, araştırılan parazit türlerine karşı güçlü antiparaziter etkinlik göstermiştir. Bu bulgular, bitkinin antiparaziter potansiyelini desteklemekle birlikte gelecekteki tibbi uygulamalar için umut vadetmektedir. Özellikle metronidazol dirençli *Trichomonas vaginalis* suşları üzerinde etkili olması, alternatif tedavi seçeneklerinin geliştirilmesi açısından önemlidir. Bu çalışma, *J. communis*'un tibbi bitki olarak kullanım potansiyelini ortaya koymakta olup, aktivite eşliğinde izolasyon ve yapı tayini ile ilgili araştırmalar için literatüre katkı sağlayacaktır.

Anahtar Kelimeler: Antiparazitik, Juniperus communis, Blastocystis, Juniperus communis, Leishmania, Plasmodium, Trichomonas



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INTRODUCTION

Malaria remains one of the greatest threats to human health. Among the Plasmodium species that cause this disease, Plasmodium falciparum (P. falciparum) and P. vivax are responsible for high mortality rates globally (1). The development of resistance to existing drugs necessitates the discovery of cost-effective and fast-acting new therapeutics (2-4). Similarly, leishmaniasis, a disease caused by *Leishmania* species, affects millions of people worldwide. The emergence of drug resistance in leishmaniasis treatment highlights the need for new drug candidates (5,6).

Trichomonas vaginalis (T. vaginalis) is a protozoan that is sexually transmitted and widely prevalent worldwide. The increasing prevalence of metronidazole-resistant strains underscores the necessity for investigating alternative treatment approaches (7).

The prevalence of *Blastocystis* ranges from 30% to 60% in developing countries. Individuals with immunosuppression and those in close contact with animals have been identified as being at particularly high risk of infection. In vitro and in vivo studies have reported its potential association with various gastrointestinal disorders and its significant role in irritable bowel syndrome (8).

Antimicrobial resistance is classified as one of the top 10 global health threats, requiring global collaboration to combat its life-threatening consequences. This situation prioritizes the discovery of new and effective antimicrobial agents. Accordingly, plant metabolites, such as terpenes, phenolic compounds, and alkaloids, have been widely studied for their antibacterial, antiviral, antifungal, and antiparasitic properties (9).

Juniperus communis (J. communis) (common juniper) is a plant historically used for medicinal and therapeutic purposes. In recent years, studies have investigated the antimicrobial, anticancer, and antiparasitic effects of different extracts of this plant (10).

This study aims to evaluate the antiparasitic activities of ethanol, methanol, chloroform, and water extracts of J. communis against various parasites, thereby exploring the potential of natural products in medical applications. The findings obtained will enhance our understanding of this plant's antiparasitic properties and guide future research.

METHODS

Ethical Approval

This research was approved by the Ethics Committee of the Faculty of Medicine, Manisa Celal Bayar University (approval date: 29/03/2023; approval number: 20.478.486/1773).

Preparation of J. communis Extracts

Ethanol, methanol, chloroform, and water extracts of J. communis fruits were obtained by the maceration technique. 5 g of ground fruits were stirred with 100 mL of solvent and macerated at room temperature for 24 hours. Each extract solution was filtered, and the solvent was evaporated under reduced pressure in a rotary evaporator until dryness. The extracts were stored at -20 °C until use.

Parasite Isolates

The parasite strains used in this study, P. falciparum (3D7), Leishmania tropica (MHOM/TR/2012/CBCL-LT), T. vaginalis (ATCC-50143), and Blastocystis, were obtained from the Parasite Bank of the Faculty of Medicine, Manisa Celal Bayar University.

In vitro Cultivation of P. falciparum and Screening of **Extracts**

The cultivation of the P. falciparum 3D7 strain was performed using a specialized medium prepared with 10.43 g of RPMI 1640, 25 mL of 1 M HEPES solution, 2 g of NaHCO₂, 0.5 mL of gentamicin solution, 0.272 g of hypoxanthine, and 5 g of albumax

A suspension containing 1% parasitemia was distributed into 96-well microplates. The microplates were placed in a chamber (microaerophilic incubation environment) with a gas mixture of $5\% \text{ CO}_2$, $5\% \text{ O}_2$, and $90\% \text{ N}_2$. The chamber was incubated at 37 °Cfor 2.5 hours.

Following incubation, plant extracts were added to the parasite suspension in the microplates at final concentrations ranging from 250 µg/mL to 2.5 µg/mL, and the plates were incubated for 6 hours. After incubation, the microplates were washed, fresh medium was added, and the plates were incubated for 66 hours. At the end of the incubation, thin smears were prepared from the microplate wells. The smears were stained with Giemsa stain and

In vitro Cultivation of Leishmania tropica and Screening of Extracts

examined under a light microscope (11-14).

The Leishmania tropica isolate MHOM/TR/2012/CBCL-LT, which was isolated in Türkiye and stored in liquid nitrogen, was thawed under appropriate conditions and cultured in Novy-MacNeal-Nicolle (NNN) medium. Promastigotes grown in the NNN medium were subsequently inoculated into RPMI-1640 (Roswell Park Memorial Institute medium 1640) medium containing 10% fetal bovine serum. The growth status of the parasites was monitored every other day after inoculation. For the experiments, L. tropica promastigotes in the logarithmic phase, reaching a density of 106 promastigotes/mL, were used.

The efficacy of J. communis extracts against L. tropica promastigotes was evaluated using the CellTiter-Glo® Luminescent Cell Viability Assay (Promega, USA) kit (15-17).

In vitro Cultivation of T. vaginalis and Screening of **Extracts**

The T. vaginalis strain ATCC-50143 (metronidazole-resistant), stored in liquid nitrogen, was thawed under appropriate conditions and cultured in a Trypticase-Yeast Extract-Maltose medium. The growth intensity of the parasites was assessed by checking the media on consecutive days. For the experiments, T. vaginalis trophozoites in the logarithmic phase, reaching a density of 104 trophozoites/mL, were used.

The efficacy of J. communis extracts against T. vaginalis was evaluated using the microdilution method (7).

In vitro Cultivation of Blastocystis and Screening of **Extracts**

The Blastocystis strain, stored in liquid nitrogen, was thawed under appropriate conditions and transferred into tubes containing modified Iscove's Dulbecco's Medium supplemented with 10% inactivated horse serum. The tubes were incubated in an anaerobic environment at 35 °C. The growth status of the parasites was monitored every 2-3 days. For the experiments, Blastocystis parasites in the logarithmic phase, reaching a density of 10⁵ parasites/mL, were used.

The efficacy of *J. communis* extracts against *Blastocystis* was evaluated using the microdilution method (18-20).

Statistical Analysis

During all invitro experiments, statistical analyses were conducted to compare the effects of different J. communis fruit extracts (ethanol, methanol, chloroform, and water) on P. falciparum, L. tropica, T. vaginalis, and Blastocystis. The efficacy of the extracts was evaluated based on IC_{50} and LD values. Data analysis was performed using the chi-square test. All statistical analyses were done using SPSS (Statistical Package for the Social Sciences) software (version 21).

RESULTS

In vitro Screening of P. falciparum for Active Extracts

Examination of Giemsa-stained preparations revealed that the chloroform extract of J. communis fruits eliminated parasites at a concentration of 15 μ g/mL. In contrast, parasites were observed in all dilutions of the other extracts.

In vitro Cultivation of Leishmania tropica and Screening of Active Extracts

The antileishmanial activities of J. communis extracts were evaluated at dilutions ranging from 250 $\mu g/mL$ to 2.5 $\mu g/mL$. The IC_{50} value of amphotericin B was determined to be 0.06 μM . The IC_{50} values for the ethanol, methanol, and water extracts of J. communis were >250 $\mu g/mL$, while the IC_{50} value for the chloroform extract was determined to be 10 $\mu g/mL$.

In vitro Cultivation of T. vaginalis and Screening of Active Extracts

The IC_{50} values representing the antitrichomonal activities of the extracts were evaluated. The IC_{50} values for the ethanol, methanol, and water extracts of J. communis fruits were >250 $\mu g/mL$, whereas the IC_{50} value for the chloroform extract was determined to be 30 $\mu g/mL$.

In vitro Cultivation of Blastocystis and Screening of Active Extracts

The activities of *J. communis* extracts against *Blastocystis* were assessed at dilutions ranging from 250 $\mu g/mL$ to 2.5 $\mu g/mL$. The lethal concentration (LD) values for the ethanol, methanol, and water extracts were found to be >250 $\mu g/mL$, whereas the LD value for the chloroform extract was determined to be 30 $\mu g/mL$.

Statistical Analysis

According to the chi-square test, a significant difference was observed between the chloroform extract and the other extracts in the *in vitro* efficacy against *P. falciparum*, *L. tropica*, *T. vaginalis*, and *Blastocystis* (p<0.05).

DISCUSSION

Juniperus species have long been used in various cultures for their antiparasitic properties (21). Similarly, the leaves and fruits of *Juniperus oxycedrus* (prickly juniper) have been applied topically to treat parasitic diseases (22). In this study, the ethanol, methanol, chloroform, and water extracts of *J. communis* were comprehensively evaluated for their *in vitro* effects on *P.*

falciparum, *L. tropica*, *T. vaginalis*, and *Blastocystis*. The results indicate that this plant exhibits antiparasitic properties.

The antimalarial potential of *Juniperus* species has been particularly investigated for *J. communis*. Essential oils derived from *J. communis* have been reported to be tested against both chloroquine-sensitive and chloroquine-resistant strains of *P. falciparum*. Analyses using Gas Chromatography and Gas Chromatography-Mass Spectrometry have shown that the essential oils contain common components such as α -pinene, although the specific components vary between oils. However, it has been reported that the essential oil of *J. communis* exhibits limited antimalarial activity, with IC_{50} values exceeding 1000 µg/ mL, indicating low efficacy in inhibiting parasite growth (23).

In our study, the chloroform extract of $\it J. communis$ fruits demonstrated significant activity against $\it P. falciparum$. The chloroform extract exhibited a completely parasiticidal effect at a concentration of 15 $\mu g/mL$. However, the low efficacy of the ethanol, methanol, and water extracts suggests that these compounds have limited antimalarial activity.

In addition, *Juniperus procera*, a close relative of *Juniperus communis*, has been reported to exhibit notable antimalarial and antileishmanial properties. The n-hexane fraction of the ethanol extract of *Juniperus procera* was found to exhibit the most prominent activity among the tested fractions. Various diterpenes, such as abieta-7,13-diene, were isolated from this fraction, and this compound demonstrated moderate antimalarial activity against *P. falciparum* D6 and W2 clones, with IC $_{50}$ values of 1.9 and 2.0 µg/mL, respectively. The IC $_{50}$ values of the crude n-hexane fraction were 5.8 and 4.4 µg/mL, suggesting potential synergistic effects among the components of the extract or the presence of additional potent antimalarial compounds (24).

More comprehensive studies on *Juniperus excelsa* have revealed its significant antileishmanial effects. A study evaluating the efficacy of Greek juniper leaf and fruit extracts against leishmaniasis reported high antileishmanial activity. The petroleum ether and chloroform fractions demonstrated particularly strong activity, showing high efficacy even at low concentrations. These findings highlight the potential of Greek juniper extracts as effective antileishmanial agents (25). In our study, the high IC $_{50}$ values of ethanol, methanol, and water extracts against $Leishmania\ tropica$ promastigotes indicate weak antileishmanial activity for these extracts. However, the chloroform extract exhibited strong activity at a concentration of 10 $\mu g/mL$.

A comprehensive study conducted in 2013 at Shiraz University of Medical Sciences evaluated the antileishmanial activities of Juniperus excelsa leaf and fruit extracts, as well as leaf fractions, against Leishmania major in both in vitro and in vivo models. The leaf extract was reported to exhibit the highest efficacy (IC_{so}: 0.97±3.53 mg/mL), and the ethyl acetate fraction showed significant activity (IC₅₀: 1.95±5.30 mg/mL). In the *in vivo* study, a significant reduction in lesion size was observed in the test group compared to the control group (p<0.05), suggesting that *Juniperus excelsa* could contribute to antileishmanial therapy (26). Additionally, a placebo-controlled clinical trial evaluated the efficacy of a topical formulation of Juniperus excelsa leaf extract for the treatment of cutaneous leishmaniasis. Seventy-two patients were treated with the extract in a placebo-controlled manner for three months, with cryotherapy administered as standard treatment. Weekly assessments showed that 82% of patients in the extract group achieved complete recovery, compared to 34%

in the placebo group, demonstrating a significant difference (p<0.001). The recovery time was also shorter in the extract group (p=0.04), with no major side effects reported other than mild local irritation in some patients. The researchers concluded that Juniperus excelsa extract is an effective adjunctive treatment for cutaneous leishmaniasis when combined with cryotherapy, increasing recovery rates and shortening recovery time (27).

In this study, the chloroform extract of *J. communis* demonstrated activity against metronidazole-resistant T. vaginalis at a concentration of 30 µg/mL, suggesting its potential as an alternative treatment for resistant strains. In contrast, the ethanol, methanol, and water extracts exhibited lower efficacy. Further investigation into the antitrichomonal properties of these extracts may help to elucidate the observed differences in activity.

The chloroform extract demonstrated lethal activity against Blastocystis at a concentration of 30 µg/mL. In contrast, the ethanol, methanol, and water extracts showed limited efficacy, even at higher concentrations, consistent with previous findings for P. falciparum, L. tropica, and T. vaginalis samples.

These findings highlight the diverse antiparasitic properties of Juniperus species, demonstrating their broad potential for medical applications. However, the effectiveness of different Juniperus species and their extracts varies against different parasites. While J. communis showed limited activity against malaria, Juniperus procera and Juniperus excelsa have shown more promising results against leishmaniasis and malaria. Further investigation of the active components in these species and the optimization of potential therapeutic applications are needed.

Future studies should focus on understanding the full spectrum of components contained within these plants and exploring the synergistic effects of these components. This will enable the development of more effective and safer antiparasitic therapies using Juniperus species.

Although our study comprehensively evaluated the antiparasitic activities of various J. communis extracts, there are some important limitations. The cytotoxic activities of the extracts have not yet been tested, which poses a limitation for their potential therapeutic applications in humans. Additionally, the efficacy of the extracts has not been tested in in vivo models, creating a gap in confirming their biological activity in complex systems and assessing their clinical potential.

CONCLUSION

This study demonstrates that the chloroform extract of J. communis exhibits strong antiparasitic activity against specific parasite species. The findings support the medicinal potential of this plant and offer promising prospects for future therapeutic applications. Notably, its effectiveness against metronidazoleresistant T. vaginalis strains is significant for the development of alternative treatment options. However, the low efficacy of ethanol, methanol, and water extracts highlights the need for more detailed investigations into their components and application methods. This study underscores the potential of *J.* communis as a medicinal plant and contributes to the literature for further research.

*Ethics

Ethics Committee Approval: This research was approved by the Ethics Committee of the Faculty of Medicine, Manisa Celal Bayar University (approval date: 29/03/2023; approval number: 20.478.486/1773).

Informed Consent: Since this study was conducted solely using archived parasite isolates in laboratory settings, informed consent was not required.

Footnotes

*Authorship Contributions

Concept: İ.Ç., Y.Ö., V.T., H.K., K.Y., A.Ö., Design: V.T., K.Y., A.Ö., Data Collection or Processing: Y.Ö., V.T., Analysis or Interpretation: H.K., K.Y., A.Ö., Literature Search: İ.Ç., Y.Ö., Writing: İ.C., A.Ö.

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REFERENCES

- 1. Rahimi BA, Thakkinstian A, White NJ, Sirivichayakul C, Dondorp AM, Chokejindachai W. Severe vivax malaria: a systematic review and metaanalysis of clinical studies since 1900. Malar J. 2014; 13: 481.
- 2. Capela R, Moreira R, Lopes F. An overview of drug resistance in protozoal diseases. Int J Mol Sci. 2019; 20: 5748.
- Nsanzabana C. Resistance to artemisinin combination therapies (ACTs): do not forget the partner drug! Trop Med Infect Dis. 2019; 4: 26.
- Pandey SK, Anand U, Siddiqui WA, Tripathi R. Drug development strategies for Malaria: with the hope for new antimalarial drug discoveryan update. Adv Med. 2023; 2023: 5060665.
- Ünübol N, Çavuş İ, Polat T, Kurt Ö, Özbilgin A, Kocagöz T. Antimicrobial peptides and their anti-leishmanial efficacies on Leishmania tropica promastigotes In vitro. Turkiye Parazitol Derg. 2024; 48: 135-41. English.
- Sundar S, More DK, Singh MK, Singh VP, Sharma S, Makharia A, et al. Failure of pentavalent antimony in visceral leishmaniasis in India: report from the center of the Indian epidemic. Clin Infect Dis. 2000; 31: 1104-7.
- Özel Y, Çavuş İ, Ünlü G, Ünlü M, Özbilgin A. Investigation of the antitrichomonal activity of cinnamaldehyde, carvacrol and thymol and synergy with metronidazole. Turkiye Parazitol Derg. 2024; 48: 72-6.
- Wawrzyniak I, Poirier P, Viscogliosi E, Dionigia M, Texier C, Delbac F, et al. Blastocystis, an unrecognized parasite: an overview of pathogenesis and diagnosis. Ther Adv Infect Dis. 2013; 1: 167-78.
- Arip M, Selvaraja M, R M, Tan LF, Leong MY, Tan PL, et al. Review on plantbased management in combating antimic robial resistance - mechanistic $% \left(1\right) =\left(1\right) \left(1\right) =\left(1\right) \left($ perspective. Front Pharmacol. 2022; 13: 879495.
- 10. Gonçalves AC, Flores-Félix JD, Coutinho P, Alves G, Silva LR. Zimbro (Juniperus communis L.) as a promising source of bioactive compounds and biomedical activities: a review on recent trends. Int J Mol Sci. 2022;
- 11. Özbilgin A, Çavuş İ, Nuraydın A, Kaya T. In vivo and in vitro models for scanning drug substances in Malaria: prestudy. Türkiye Parazitol Derg. 2017; 41: 156-63.
- 12. Witkowski B, Amaratunga C, Khim N, Sreng S, Chim P, Kim S, et al. Novel phenotypic assays for the detection of artemisinin-resistant *Plasmodium* falciparum malaria in Cambodia: in-vitro and ex-vivo drug-response studies. Lancet Infect Dis. 2013; 13: 1043-9.

- Amaratunga C, Lim P, Suon S, Sreng S, Mao S, Sopha C, et al. Dihydroartemisinin-piperaquine resistance in *Plasmodium falciparum* malaria in Cambodia: a multisite prospective cohort study. Lancet Infect Dis. 2016; 16: 357-65.
- 14. Witkowski B, Menard D, Amaratunga C, Fairhurst RM. Ring stage survival assays (RSA) to evaluate the *in vitro* and *ex vivo* susceptibility of *Plasmodium falciparum* to artemisinins. National Institutes of Health Procedure RSAv1. 2013; 1-16.
- 15. Güler E, Özbilgin A, Çavuş İ, Baddal B, Etikan İ, Başer KHC, et al. *In vitro* anti-leishmanial activity of essential oils extracted from plants growing in Northern Cyprus against *Leishmania tropica*. Turkiye Parazitol Derg. 2021; 45: 101-7. English.
- Özbilgin A, Çavuş İ, Yıldırım A, Kaya T, Ertabaklar H. Evaluation of in vitro and in vivo drug efficacy over leishmania tropica: a pilot study. Turkiye Parazitol Derg. 2018; 42: 11-9.
- Eser M, Çavuş İ. In vitro and in silico evaluations of the antileishmanial activities of new benzimidazole-triazole derivatives. Vet Sci. 2023; 10: 648.
- Mirza H, Teo JD, Upcroft J, Tan KS. A rapid, high-throughput viability assay for *Blastocystis* spp. reveals metronidazole resistance and extensive subtype-dependent variations in drug susceptibilities. Antimicrob Agents Chemother. 2011; 55: 637-48.
- Mei X, Wei L, Su C, Yang Z, Tian X, Zhang Z, et al. Advances in the axenic isolation methods of *Blastocystis sp.* and their applications. Parasitology. 2024; 151: 125-34.
- Mokhtar AB, Ahmed SA, Eltamany EE, Karanis P. Anti-Blastocystis activity in vitro of Egyptian herbal extracts (family: asteraceae) with emphasis on Artemisia judaica. Int J Environ Res Public Health. 2019; 16: 1555.

- Yesilada E, Honda G, Sezik E, Tabata M, Goto K, Ikeshiro Y. Traditional medicine in Turkey. IV. Folk medicine in the Mediterranean subdivision. J Ethnopharmacol. 1993; 39: 31-8.
- Sezik E, Yesilada E, Tabata M, Honda G, Takahashi Y, Fujita T, et al. Traditional medicine in Turkey. VIII. Folk medicine in East Anatolia: Erzurum, Erzincan, Agri, Kars, Igdir Provinces. Econ Bot. 1997; 51: 195-211
- Milhau G, Valentin A, Benoit F, Mallié M, Bastide JM, Pélissier Y, et al. In vitro antimalarial activity of eight essential oils. J Essent Oil Res. 1997; 9: 329-33.
- Samoylenko V, Dunbar DC, Gafur MA, Khan SI, Ross SA, Mossa JS, et al. Antiparasitic, nematicidal and antifouling constituents from Juniperus berries. Phytother Res. 2008; 22: 1570-6.
- Moein M, Hatam G, Taghavi-Moghadam R, Zarshenas MM. Antileishmanial activities of Greek juniper (*Juniperus excelsa* M.Bieb.) against *Leishmania major* promastigotes. J Evid Based Complement Alternat Med. 2017; 22: 31-6.
- 26. Mirzavand S, Hatam G, Moein M, Zarshenas MM. In vitro and in vivo assessment of anti-leishmanial efficacy of leaf, fruit, and fractions of Juniperus excelsa against axenic amastigotes of Leishmania major and topical formulation in Balb/c mice. Iran Red Crescent Med J. 2019; 21: 1-7.
- 27. Parvizi MM, Handjani F, Moein M, Hatam G, Nimrouzi M, Hassanzadeh J, et al. Efficacy of cryotherapy plus topical *Juniperus excelsa* M.Bieb cream versus cryotherapy plus placebo in the treatment of old world cutaneous leishmaniasis: a triple-blind randomized controlled clinical trial. PLoS Negl Trop Dis. 2017; 11: e0005957.