

# Age-Dependent Variations in the Venom Proteins of *Vipera xanthina* (Gray, 1849) (Ophidia: Viperidae)

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**SUMMARY:** In this study, polyacrylamide disc gel electrophoresis and densitometry analysis methods were used to analyze venom extracts of *Vipera xanthina* specimens of different lengths (35, 47 and 88 cm) collected from the same locality. The electropherograms of the venom protein samples showed age-dependent qualitative and quantitative variations.

**Key Words:** *Vipera xanthina*, Venom proteins, Polyacrylamide gel electrophoresis, Densitometry

## *Vipera xanthina* (Gray, 1849) (Ophidia: Viperidae) Türünün Venom Proteinlerinde Yaşa Bağlı Varyasyonlar

**ÖZET:** Mevcut çalışmada aynı lokaliteden (Gümüldür-İzmir) toplanan üç farklı boyda (35 cm, 47 cm and 88 cm) *Vipera xanthina*'nın venom ekstraktları poliakrilamid jel elektroforezi ve densitometri yöntemleri ile analiz edilmiştir. İncelenen venom protein örneklerinin elektroferogramları arasında yaşa bağlı kalitatif ve kantitatif varyasyonlar tespit edilmiştir.

**Anahtar Sözcükler:** *Vipera xanthina*, Zehir proteinleri, Poliakrilamid jel elektroforezi, Densitometri

## INTRODUCTION

The Ottoman Viper, *Vipera xanthina* (Gray, 1849), an endemic species in Turkey, is known from Middle, South and West Turkey with vertical distribution to 2000 m (4). Age-dependent variations in biological features of venom extracts have been frequently reported by various researchers (10, 14, 15, 19, 20).

Studies related to snakes in Turkey have tended to deal with the taxonomy and distribution of species (3, 4, 5, 7, 21). Recently, studies were carried out on the pattern of the venom proteins of different colubrid and viperid species inhabiting Turkey (1, 2).

This study examines venom proteins of *Vipera xanthina* specimens collected from the same locality. All venom extracts, obtained from three different specimens having different lengths, were analyzed with polyacrylamide disc gel electrophoresis, and their venom electrophoretic patterns were compared.

## MATERIAL AND METHODS

Specimens used in this study were collected in the same locality (Gümüldür, İzmir 42°14'N - 27°02'E). The specimens were taken to the laboratory alive and total lengths were measured using a dial caliper with accuracy  $\pm 0.02$  mm. Thereafter, venom was extracted without applying any pressure on the venom glands, as described by Tare et al. (18). As venom extracts also contain dead cells, they were centrifuged for 5 minutes at 600 g and stored in equal amounts (4  $\mu$ l venom per sample) for each separation until they were analyzed. The venom proteins were separated according to methods given by Davis (8), slightly modified by Özeti and Atatür (17). Accordingly, a pH 6.7 stacking gel was layered above the pH 9 separation gels of 7.5% polyacrylamide, together with pH 8.3 tris-glycine buffer system. Electrophoretic separations were carried out at room temperature (approx. 20 – 25 °C), utilizing a Canalco Model 1200 electrophoresis apparatus. Separation gels were stained with 0.5 % Amido Black (Naphthol Blue Black 10-B), later destained passively with repeated 7 % acetic acid baths. Then, the stained gels were photographed. Qualitative evaluation of the gels was done directly from the electropherograms, and the densitometric curves of the separations were obtained by means of a Gelman ACD-15 Model 39430 densitometer at 500 nm.

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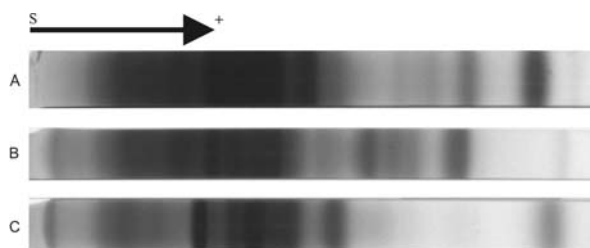
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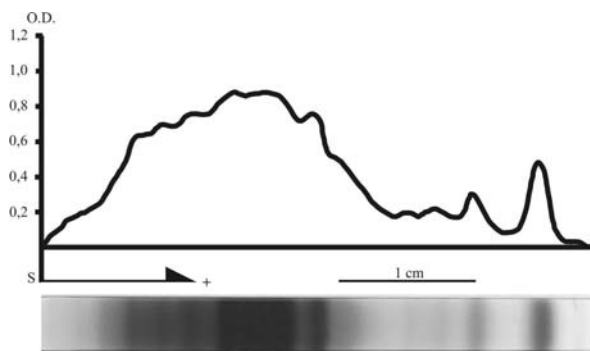
## RESULTS

The examined venom secretions of *Vipera xanthina* had a higher viscosity than that of the water. Although venom secretions of two specimens (47 and 88 cm total lengths) were light yellow, they were colorless in the 35-cm-long specimen.

The gel photographs of the venom protein samples are given in Fig. 1. According to the electrophoretic analyses, the proteins could be separated into 13 fractions or fraction groups in the two shorter specimens (35 and 47 cm), while 14 fractions were recorded in the longest individual (88 cm). Comparing the electropherograms of the three specimens revealed considerable quantitative differences in terms of the venom protein fractions of the two shorter specimens, but they were similar in the number of fractions and electrophoretic mobilities. The 88 cm individual, however, differed strongly from the other two in the number of fractions, electrophoretic mobilities and densities of the venom proteins (Figs. 1-4).



**Figure 1.** Polyacrylamid gel electrophoresis of venoms of snakes of different lengths. **A.** 35 cm, **B.** 47 cm, **C.** 88 cm (S: Start, junction between the stacking and separation gels).

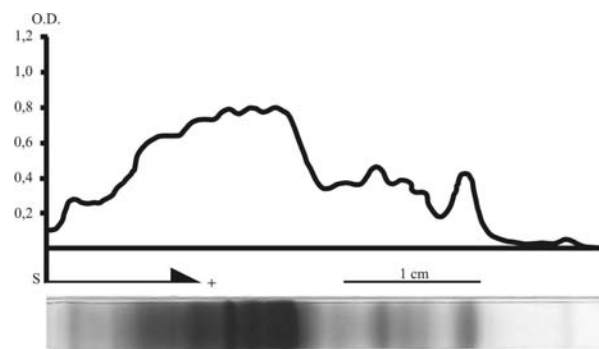


**Figure 2.** Gel photograph showing the electrophoretic separation of the venom protein sample obtained from the 35 cm length of *V. xanthina* specimen, together with its densitometric tracing curve (O.D.: Optical density, S: Start, junction between the stacking and separation gels).

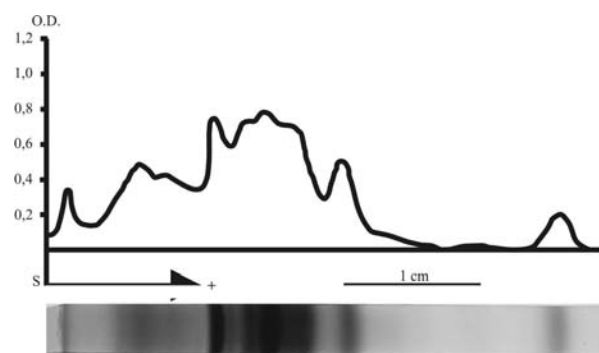
## DISCUSSION

Various studies on venom proteins of different snakes show that younger individuals have a higher potential than older ones with regard to certain activities such as lethality, coagulation and defibrination (6, 9, 10, 11, 14, 16, 19, 20).

Tun-Pe et al. (20) studied Russell's viper *Daboia russelli siamensis* venoms using SDS-PAGE electrophoresis in different-sized



**Figure 3.** Gel photograph showing the electrophoretic separation of the venom protein sample obtained from the 47 cm length of *V. xanthina* specimen, together with its densitometric tracing curve. For further explanation, see legend to fig. 2.



**Figure 4.** Gel photograph showing the electrophoretic separation of the venom protein sample obtained from the 88 cm length of *V. xanthina* specimen, together with its densitometric tracing curve. For further explanation, see legend to fig. 2.

specimens. The venom extracts of that species are colorless and have a lower activation of L-amino acid oxides. The authors also recorded the maximum activities in the venom of 40-60 cm long snakes and reported that these activities decreased at older snakes depending on length and age. Similar results were found in the early studies on venom proteins of various other snake species (6, 10, 11, 12, 19).

A study on the venom proteins of the Common Lancehead (*Bothrops atrox* Linnaeus, 1758) showed that youngest snakes have fewer protein bands; the number of bands increased as the snakes aged (13), corroborating the results of Tun-Pe et al (20).

Comparing our results on the color of the venom proteins with literature data, *Vipera xanthina* is similar to other viperid species (1, 2, 20).

Among the *Vipera xanthina* specimens we examined, the most aggressive behaviour was observed in the 47-cm-long snake, while the most submissive individual was the 88-cm-long one. This study indicated that the fractions and concentration of the venom proteins are higher in the older than in the youngest specimen. Our results support those of Meier (13) and Tun-Pe et al. (20) on venom proteins of different snake species.

## REFERENCES

1. Arıkan H, Kumlutaş Y, Türkozan O, Baran İ, 2003. Electrophoretic Patterns of Some Viper Venoms from Turkey. *Turk J Zool*, 27: 239-242.
2. Arıkan H, Göçmen B, Mermer A, Bahar H, 2005. An electrophoretic comparison of the venoms of a colubrid and various viperid snakes from Turkey and Cyprus, with some taxonomic and phylogenetic implications. *Zootaxa*, 1038: 1-10.
3. Baran İ, 1976. *Türkiye yılanlarının taksonomik revizyonu ve coğrafik dağılışı*. TÜBİTAK Yayınları, No: 309, Ankara, 177 pp.
4. Baran İ, Atatür MK, 1998. *Turkish Herpetofauna (Amphibians & Reptiles)*. Republic of Turkey Ministry of Environment, Ankara, 214 pp.
5. Başoğlu M, Baran İ, 1980. *Türkiye Sürüngenleri*, Kısım II, Yılanlar (Turkish Reptiles, Part II, Snakes). Ege Üniversitesi Kitaplar Serisi No: 81, Bornova-İzmir, 218 pp.
6. Bonilla CA, Horner NV, 1969. Comparative electrophoresis of *Crotalus* and *Agkistrodon* venoms from North American snakes. *Toxicon*, 7: 327-329.
7. Budak A, Göçmen B, 2005. Herpetoloji. E. Ü. Yayınları, Fen Fakültesi Yayın No:194, E. Ü. Basımevi, 230 sayfa.
8. Davis BJ, 1964. Disc Electrophoresis-II, Method and Application to Human Serum Proteins. *Annals of New York Academy Sciences*, 121: 404-427.
9. Fiero MK, Siefert MW, Weaver TJ, Bonilla CA, 1972. Comparative study of juvenile and adult prairie rattlesnake (*Crotalus viridis viridis*) venoms. *Toxicon*, 10: 81-82.
10. Furtado MFD, Maruyama M, Kamuguti AS, Antonio LC, 1991. Comparative study of nine *Bothrops* snake venoms from adult female snakes and their offspring. *Toxicon*, 29: 219-226.
11. Glenn JL, Straight RC, 1985. Venom properties of the rattlesnakes (*Crotalus*) inhabiting the Baja California region of Mexico. *Toxicon*, 23: 769-775.
12. Jimenez-Porras JM, 1964. Intraspecific variation of venom of the jumping viper *Bothrops nummifera*. *Toxicon*, 2: 187-195.
13. Meier J, 1986. Individual and age-dependent variations in the venom of the Fer-de-lance (*Bothrops atrox*). *Toxicon*, 24 (1): 41-45.
14. Meier J, Freyvogel TA, 1980. Comparative studies on venoms of the fer-de-lance (*Bothrops atrox*), carpet viper (*Echis carinatus*) and spitting cobra (*Naja nigricollis*) snakes at different ages. *Toxicon*, 18: 661-662.
15. Minton SA, 1967. Observations on toxicity and antigenic makeup of venoms from juvenile snakes. In: Russell, F. E. and Saunders, P. R. (Ed), Animal Toxins, pp. 211-222, Pergamon Press, Oxford.
16. Minton SA, 1975. A note on the venom of an aged rattlesnake. *Toxicon*, 13: 73-74.
17. Özeti N, Atatür MK, 1979. A preliminary survey of the serum proteins of a population of *Mertensiella luschani finikensis* Basoğlu&Atatür from Finike in Southwestern Anatolia. *Istanbul Üniversitesi Fen Fakültesi Mecmuası*, 44B: 23-29.
18. Tare TG, Sutar NK, Renapurkar DM, 1986. A study of snake venom yield by different methods of venom extraction. *Amphibia-Reptilia*, 7: 187-191.
19. Theakston RDG, Reid HA, 1978. Changes in the biological properties of venom from *Crotalus atrox* with aging. *Period. Biol.*, 80 (Suppl. 1): 123-133.
20. Tun-Pe, Nu-Nu-Lwin, Aye-Aye-Myint, Kyi-May-Htwe, Aung-Cho, 1995. Biochemical and biological properties of the venom from Russell's Viper (*Daboia russelli siamensis*) of varying ages. *Toxicon*, 33: 817-821.
21. Ugurtaş İH, Papenfuss TJ, Orlov NL, 2001. New record of *Walterinnesia aegyptia* Lataste, 1887 (Ophidia: Elapidae: Bungarinae) in Turkey. *Russian J Herpetol*, 8 (3): 239-245.